

Association Studies of Transforming Growth Factor- β 1 and Alzheimer's Disease

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Substantial laboratory evidence suggests Transforming Growth Factor- β 1 (*TGF β 1*) is linked to Alzheimer's Disease (AD) pathology. The purpose of the study was to estimate the genetic association of *TGF β 1* with AD while controlling for apolipoprotein E4 allele (*APOE4*) status, the only well-established genetic risk factor for AD. Two study populations were genotyped for the *TGF β 1*-509 and +869 single nucleotide polymorphisms (SNPs) that have been associated with *TGF β 1* levels. Constituting these populations were 203 families from the NIMH AD Genetic Initiative with at least two affected siblings and a normal sibling, and a population of 126 African-American (AA) AD cases versus 93 age matched controls. Results from family-based analyses showed a significant association between the *TGF β 1* -509 SNP and AD for the entire set of 203 families ($P = 0.007$), and a subset of these families without a homozygous *APOE4* family member ($P = 0.026$). Results from family-based analyses on the *TGF β 1* +869 SNP were not significant in the 203 families. While associations for the main effects of the *TGF β 1* +869 and -509 SNP with AD in the AA case-control study were also not significant, results did indicate that *TGF β 1* may function as an effect modifier of *APOE4* risk. Specifically, the odds of AD associated with having at least one *APOE4* allele increased in an additive fashion with one or two copies of the higher producer allele when stratified by *TGF β 1* -509 genotype and by *TGF β 1* +869 genotype. Results support a role for *TGF β 1* in AD pathogenesis. © 2005 Wiley-Liss, Inc.

KEY WORDS: cytokines; neuroinflammation; apolipoprotein; *TGF β 1* +869 SNP; *TGF β 1* -509 SNP

INTRODUCTION

Alzheimer's Disease (AD) is a neurodegenerative disorder with a complex etiology and pathogenesis. It is characterized by progressive memory loss and depletion of cognitive functions. The biological processes stimulating the development of AD pathology are not completely understood. One characteristic of neuropathological autopsy findings is the presence of

extracellular plaques consisting of aggregates of amyloid- β ($A\beta$) peptides, which are the proteolytic cleavage products of the amyloid- β precursor protein (APP) [Neve et al., 2000]. The function of APP is not confirmed, but APP is speculated to support formation and maintenance of synapses in vivo [Mattson et al., 1997; Wang et al., 2005].

A genetic component of AD has long been recognized, since clustering of this disease within families has been observed [Kamboh, 2004]. The only well-established genetic risk factor for AD is the E4 allele of the apolipoprotein (*APOE*) gene. The exact role of the *APOE* protein in AD pathology is not clear, however it is known that *APOE* functions as a cholesterol transporter, where *APOE* rich lipoprotein complexes are one source of cholesterol for membrane synthesis and maintenance in the brain [Lahiri et al., 2004; Poirier, 2005]. These complexes also bind $A\beta$, however, the *APOE4* isoform does not form complexes with $A\beta$ peptides, as well as other two *APOE* isoforms (*APOE2* and *APOE3*), therefore *APOE4* may result in lower efficiency in $A\beta$ clearance [LaDu et al., 1997]. Several studies report AD patients homozygous for the E4 allele account for a fraction (<30%) of all AD cases [Kamboh, 2004].

It is well established that *APOE4* is only predictive of AD onset and its influence varies across populations [Kamboh, 2004]. Therefore, it remains necessary to consider other genes that might influence the development of disease. Transforming Growth Factor- β 1 (*TGF β 1*) is a multifunctional cytokine with pro- and anti-inflammatory properties and, in particular, is a key regulator of the brain's responses to injury and inflammation [Mattson et al., 1997; Akiyama et al., 2000]. Within the CNS, *TGF β 1* is expressed in neurons, astrocytes, and microglia [Mattson et al., 1997; Harris-White et al., 1998]. Studies show increased *TGF β 1* levels in the cerebral spinal fluid of AD patients versus controls, and *TGF β 1* is detected in senile plaques [van der Wal et al., 1993; Zetterberg et al., 2004].

Two single nucleotide polymorphisms (SNPs) located in *TGF β 1* have been shown to be related to *TGF β 1* levels. The T allele of the -509 SNP located in the promoter has been associated with higher plasma concentrations of *TGF β 1* in twins, in a dose-dependent fashion [Grainger et al., 1999]. In addition, Luedeking et al. [2000] reported that the T allele was marginally associated with higher transcription activity than the other allele and that the frequency of the TT genotype was significantly higher in AD patients as compared to controls. The +869 SNP is located in the signal peptide region and results in a leucine \rightarrow proline substitution affecting the protein's polarity and its export, which could affect protein production [Wood et al., 2000]. Both a clinical osteoporosis study and a hypertension study found associations between the +869 C SNP and higher *TGF β 1* levels, thus implying that the proline (C) substitution could lead to increased *TGF β 1* production [Yamada et al., 1998; Suthanthiran et al., 2000].

In order to investigate the association of *TGF β 1* with AD, the -509 C \rightarrow T promoter SNP and the +869 T \rightarrow C SNP in the signal peptide region were genotyped in a cohort of 203 families with at least two AD affected siblings and one unaffected sibling from the NIMH AD Genetics Initiative sibling dataset [Blacker et al., 2003] and in an expanded Alabama African-American (AA) case-control cohort (N = 219) [Perry et al.,

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2001]. The purpose of this study is to estimate the effects of TGFB1 SNPs on AD risk singly and in the context of APOE4 exposure.

MATERIALS AND METHODS

Study Population

Comprising the family study group were 203 families with at least two AD affected siblings (onset ≥50 years) with mean age of onset (MAO) 71.0 ± 7.5 years and one unaffected sibling. This group (candidate gene set) is a subset of a larger group of families collected as part of the NIMH AD Genetics Initiative [Blacker et al., 2003] following a standardized protocol utilizing the NINCDS-ADRDA criteria for diagnosis of definite and probable AD [Tierney et al., 1988]. Since parental genotypes are not available for this dataset, a subset of families with two affected siblings and at least one unaffected sibling were necessary for family-based statistical analysis. The candidate gene set was further subdivided into 64 APOE4/APOE4 (E4) families with at least one member who is homozygous for the E4 allele (MAO = 67.6 ± 7.1 years), and 139 non-homozygous E4 (NHE4) families where no individuals possess the E4/E4 genotype (MAO = 72.3 ± 7.1 years). Stratifying on homozygous APOE4 status rather than carrier status separates families with strong APOE4 presence from those with no or weak APOE4 presence.

Comprising the AA case-control group were 126 AD cases and 93 age and race matched controls previously reported on by Perry et al. [2001]. AA AD patients had a MAO of 70.8 ± 7.8 years, while the mean age of the control group was 75.2 ± 7.6 years. Cases were identified from patients seen in the Memory Disorder Clinic at the University of Alabama at Birmingham utilizing the NINCDS-ADRDA criteria mentioned above for diagnosis of definite and probable AD. Additional neurological and laboratory tests were completed to exclude cases of other illnesses that can mimic AD. Unaffected spouses of AD patients and members of AD support groups, who screened negative for a personal and family history of memory problems and scored normal on cognitive screening, comprised the control group.

Determination of APOE and TGFB1 Genotypes

Blood from the NIMH Genetics Initiative families was collected and sent to the NIMH repository at Rutgers where genomic DNA was extracted from lymphocyte cell lines. Isolation of genomic DNA from the AA subjects has been described [Perry et al., 2001].

APOE4 genotyping followed a modified version of the procedure from Hixson and Vernier [1990] [Blacker et al., 1997]. Primers and enzymes for the PCR-RFLP analysis of the -509 and +869 SNPs of TGFB1 were obtained from Luedeking et al. [2000] and Wood et al. [2000], respectively. All PCRs were performed on MJ thermocyclers (Watertown, MA). Amplification of both SNPs were performed in 25 µl reactions containing 0.5 µM of each primer, 1.5 µM MgCl2, 200 µM of each dNTP, 1U Taq (Promega, Madison, WI) and 100 ng of DNA. Conditions were 30 cycles of 30 sec cycling times, with the annealing temperatures for the -509 and +869 SNPs set at 64 and 58°C, respectively. The microtiter plates used in the PCRs were allowed to cool at 12°C/30 min to reduce condensation. Digestions of the -509 and +869 products were performed with 2 and 3U of enzyme (N.E. Biolabs, Beverly, MA), respectively in 15 µl reactions containing 8 µl of product at 37°C/4 hr. Digested products were separated on 2% SFR agarose gels (Amresco, Solon, OH) and photographed on a Fluor-S imager (Bio-Rad, Hercules, CA). Two independent readers were used for assigning genotypes to reduce errors and clarify any ambiguities.

Statistical Analysis

The family-based association tests implemented in FBAT [Horvath et al., 2001] were used to examine the association of the TGFB1 SNPs with AD in the candidate gene family set and the E4 and NHE4 subsets. APOE4 allele dose was controlled for in FBAT in the total candidate gene set by utilizing a difference residual outcome variable from the logistic regression of APOE4 status on AD [Lunetta et al., 2000].

For the AA population, the case and control groups were tested for Hardy-Weinberg equilibrium (HWE) at the TGFB1 and APOE loci using the chi-square test. Two sample tests for binomial proportions were used to compare genotype frequencies between cases and controls for each locus. Logistic regression was used to investigate the main effects association of the TGFB1 SNPs with AD controlling for APOE4 genotype. Mantel-Haenszel methods were applied to examine the association between APOE4 and AD stratified by TGFB1 SNP genotype to further explore the possibility of an interaction between APOE4 and TGFB1 that affects disease status [Mantel and Haenszel, 1959].

RESULTS

Results of the family-based association analyses for the -509 (C → T) polymorphism of TGFB1 show a significant association (P = 0.007) with AD for the T allele in the candidate gene set, and also marginal evidence for an association in the NHE4 subset (P = 0.026). When controlling for the presence of at least one APOE4 allele in the candidate gene set in FBAT, the association between AD and the -509 SNP remained highly significant (P = 0.007). This association shows the T allele of the -509 (C → T) polymorphism of TGFB1 is transmitted more often to the affected siblings. Results from family-based analyses on the TGFB1 +869 SNP did not reveal a significant association with AD (data not shown).

The AA case-control population is in HWE for the APOE locus and the TGFB1 SNPs. Results of baseline comparisons of APOE genotype and the TGFB1 genotypes in cases compared to controls show the frequency of the APOE4 genotype differs among cases and controls (P < 0.013) as expected, but the TGFB1 genotypes do not (data not shown). The main effects models testing the association of the -509 SNP and +869 SNP genotypes with AD controlling for APOE4 status do not show an association between TGFB1 and AD. Analysis of the association of the TGFB1 -509 genotype with AD stratified by APOE4 status (having 0, 1, or 2 APOE4 alleles) also showed no significant results, as did the same analysis for the TGFB1 +869 SNP (data not shown).

The association of at least one APOE4 allele with AD stratified by TGFB1 -509 or +869 SNP genotype is displayed in Table I. Results suggest the T allele of the -509 SNP and C

TABLE I. Test for the Association of at Least One APOE4 Allele With AD Stratified by the TGFB1 -509 or +869 SNP Genotypes in the African-American Case-Control Population

| | OR | 95% CI |
|-----------------------------|------|-------------|
| TGFB1 -509 SNP | | |
| CC (n = 115) | 2.77 | 1.28, 6.01 |
| CT (n = 78) | 4.02 | 1.57, 10.33 |
| TT (n = 17) | 6.00 | 0.52, 69.75 |
| Mantel-Haenszel combined OR | 3.35 | 1.88, 5.98 |
| TGFB1 +869 SNP | | |
| TT (n = 64) | 2.23 | 0.77, 6.45 |
| CT (n = 92) | 3.53 | 1.49, 8.33 |
| CC (n = 40) | 7.43 | 1.77, 31.04 |
| Mantel-Haenszel combined OR | 3.50 | 1.93, 6.36 |

allele of the +869 SNP may modify the effects of APOE4 in an additive manner since the odds ratio between AD occurrence and presence of at least one *APOE4* allele increase with each additional copy of the T or C allele. However, these results are only suggestive since the confidence intervals for the odds ratios in the three strata of both SNPs are wide and overlap.

DISCUSSION

A significant association between the -509 *TGFB1* SNP and AD was found for the candidate gene set using the family-based association test implemented in FBAT ($P = 0.007$). The significant association indicates higher transmission of the -509 T allele to affected siblings as compared to unaffected siblings. When controlling for at least one *APOE4* allele in FBAT, the association between the *TGFB1* -509 SNP and AD remained highly significant. The *TGFB1* -509 SNP lies in a negative regulatory element of its promoter, but the T substitution appears to be associated with higher transcription activity and higher plasma levels of TGFB1 [Grainger et al., 1999; Luedeking et al., 2000]. Our results are similar to a Pittsburgh study that found the TT genotype was significantly higher in AD patients versus controls [Luedeking et al., 2000]. However, a larger group of French sporadic patients as compared to age-matched controls [Araria-Goumidi et al., 2002] failed to replicate this association.

Like the *TGFB1* -509 T substitution, the +869 C substitution may also affect TGFB1 production [Wood et al., 2000]; associations of the +869 C substitution with higher TGFB1 levels have been reported [Yamada et al., 1998; Suthanthiran et al., 2000], however, our results from family-based analyses on the +869 SNP with AD failed to find any association.

For the case-control study in the AA population, there were no associations between the *TGFB1* SNPs and AD. However, the association between carrying at least one *APOE4* allele and AD increased when stratified by the *TGFB1* -509 or the +869 SNP genotype. The odds of AD, as determined by *APOE4* carrier status, increase in an additive manner based on the number of *TGFB1* -509 T alleles or +869 C alleles (0, 1, or 2) (Table I). Although these differences are not significant due to the wide confidence intervals, it is notable that all six participants who were homozygous for *APOE4* and the *TGFB1* +869 C SNP have AD. There are no subjects homozygous for both *APOE4* and the *TGFB1* -509 T SNP. Interestingly, these results also show that having no *TGFB1* +869 C alleles dilutes the risk for AD associated with *APOE4* carrier status. The above results for the additive effects from the -509 T allele and the +869 C allele in the presence of at least one *APOE4* allele lend support for the possibility that TGFB1 levels may modify the effects of APOE4 protein in AD. Although small sample size restricts the power of these stratified analyses, the consistency of the results between these two SNPs suggests further investigation of a possible synergism between APOE4 and TGFB1 is warranted.

A model of AD that includes TGFB1 has biologic support. TGFB1 increases APP mRNA and protein expression in brain cell cultures [Gray and Patel, 1993; Monning et al., 1994] and appears to increase A β deposition in brain cell cultures and in APP and *TGFB1* transgenic mice, when overexpressed [Harris-White et al., 1998; Buckwalter et al., 2002; Lesne et al., 2003]. In addition, one transgenic mouse study showed astroglial overexpression of TGFB1 produced a strong upmodulation of extracellular matrix (ECM) proteins in the central nervous system (CNS) [Wyss-Coray et al., 1995]. ECM proteins have been identified in A β plaques of AD brains where they appear to play a central role in plaque deposition and stabilization [Fillit and Leveugle, 1995]. This model of the contribution of TGFB1 to plaque formation and stabilization is also consistent with the model in atherosclerosis, another

risk factor for AD [Forrester, 2004]. The bulk of severe clinical manifestations of atherosclerosis is attributed to disruption of plaques rich in inflammatory macrophages and T cells, that are characterized by a thin fibrous cap with substantial loss in ECM [Libby, 2001]. Transgenic mice with T cell specific inhibition of TGFB1 display plaques with these vulnerable properties of plaque T cell infiltration and loss of ECM production [Gojova et al., 2003].

How does APOE4 fit into this picture? Exposure of cultured neurons to A β peptides causes an increase in oxyradical formation and subsequent radical mediated damage to neuron membrane lipids and proteins [Markesbery, 1999]; and the APOE4 isoform has been shown to provide cultured neurons the least antioxidant protection from A β generated hydrogen peroxide compared to APOE2 or APOE3 [Miyata and Smith, 1996]. Furthermore, the *TGFB1* promoter contains an element responsive to the transcription factor AP-1 that can be regulated by redox reactions [Iglesias-De La Cruz et al., 2001; Wilmer et al., 2002]. If TGFB1 upregulates APP and A β production and deposition, and A β induces ROS production, which then upregulates TGFB1 further, the reduced antioxidant efficiency of APOE4 may lead to exacerbation of the AD pathological process. Given this model, it is plausible that TGFB1 may modify the risk for AD associated with the APOE4 isoform by cooperating with A β in aggravating the inflammatory process such that it overwhelms the lesser antioxidant capacity of APOE4.

Our results suggest that TGFB1 may be associated with AD independently as results from family-based tests indicate or it may function as an effect modifier of APOE4 risk by contributing to a chronic, inflammatory neuropathologic state as results from the case-control association analyses suggest. Biologically both models are feasible, however, both our samples were underpowered to confirm either model. Therefore, we recommend that *APOE* and *TGFB1* SNPs should be genotyped in family and case/control populations using large sample sizes with sufficient power to distinguish between these two models to confirm our suggestive findings. If either model can be substantiated for TGFB1 in AD, then the discovery of its exact role in this pathologic process from future cell specific laboratory studies and molecular genetic studies, may provide better insight into the role of cytokines in the neuroinflammatory processes underlying AD pathology, and hence lead to new targets for therapy.

REFERENCES

- Akiyama H, Barger S, Barnum S, Bradt B, Bauer J, Cole GM, Cooper NR, Eikelenboom P, Emmerling M, Fiebich BL, et al. 2000. Inflammation and Alzheimer's disease. *Neurobiol Aging* 21(3):383-421.
- Araria-Goumidi L, Lambert JC, Mann DM, Lendon C, Frigard B, Iwatsubo T, Cotel D, Amouyel P, Chartier-Harlin MC. 2002. Association study of three polymorphisms of TGF-beta1 gene with Alzheimer's disease. *J Neurol Neurosurg Psychiatry* 73(1):62-64.
- Blacker D, Haines JL, Rodes H, Terwedow H, Go RCP, Harrell LE, Perry RT, Bassett SS, Chase GA, Meyers D, et al. 1997. ApoE-4 and age at onset of Alzheimer's Disease: The NIMH Genetic Initiative. *Neurology* 48:139-147.
- Blacker D, Bertram L, Saunders AJ, Moscarillo TJ, Albert MS, Wiener H, Perry RT, Collins JS, Harrell LE, Go RC, et al. 2003. Results of a high-resolution genome screen of 437 Alzheimer's disease families. *Hum Mol Genet* 12(1):23-32.
- Buckwalter M, Pepper JP, Gaertner RF, Von Euw D, Lacombe P, Wyss-Coray T. 2002. Molecular and functional dissection of TGF-beta1-induced cerebrovascular abnormalities in transgenic mice. *Ann NY Acad Sci* 977:87-95.
- Fillit H, Leveugle B. 1995. Disorders of the extracellular matrix and the pathogenesis of senile dementia of the Alzheimer's type. *Lab Invest* 72(3):249-253.
- Forrester JS. 2004. Common ancestors: Chronic progressive diseases have the same pathogenesis. *Clin Cardiol* 27(4):186-190.

- Gojova A, Brun V, Esposito B, Cottrez F, Gourdy P, Ardouin P, Tedgui A, Mallat Z, Groux H. 2003. Specific abrogation of transforming growth factor-beta signaling in T cells alters atherosclerotic lesion size and composition in mice. *Blood* 102(12):4052-4058.
- Grainger DJ, Heathcote K, Chiano M, Snieder H, Kemp PR, Metcalfe JC, Carter ND, Spector TD. 1999. Genetic control of the circulating concentration of transforming growth factor type beta1. *Hum Mol Genet* 8(1):93-97.
- Gray CW, Patel AJ. 1993. Regulation of beta-amyloid precursor protein isoform mRNAs by transforming growth factor-beta 1 and interleukin-1 beta in astrocytes. *Brain Res Mol Brain Res* 19(3):251-256.
- Harris-White ME, Chu T, Balverde Z, Sigel JJ, Flanders KC, Frautschy SA. 1998. Effects of transforming growth factor-beta (isoforms 1-3) on amyloid-beta deposition, inflammation, and cell targeting in organotypic hippocampal slice cultures. *J Neurosci* 18(24):10366-10374.
- Hixson JE, Vernier DT. 1990. Restriction isotyping of human apolipoprotein E by gene amplification and cleavage with HhaI. *J Lipid Res* 31(3):545-548.
- Horvath S, Wei E, Xu X, Palmer LJ, Baur M. 2001. Family-based association test method: age of onset traits and covariates. *Genet Epidemiol* 21(Suppl 1):S403-S408.
- Iglesias-De La Cruz MC, Ruiz-Torres P, Alcamí J, Díez-Marques L, Ortega-Velazquez R, Chen S, Rodriguez-Puyol M, Ziyadeh FN, Rodriguez-Puyol D. 2001. Hydrogen peroxide increases extracellular matrix mRNA through TGF-beta in human mesangial cells. *Kidney Int* 59(1):87-95.
- Kamboh MI. 2004. Molecular genetics of late-onset Alzheimer's disease. *Ann Hum Genet* 68(Pt 4):381-404.
- LaDu MJ, Lukens JR, Reardon CA, Getz GS. 1997. Association of human, rat, and rabbit apolipoprotein E with beta-amyloid. *J Neurosci Res* 49(1):9-18.
- Lahiri DK, Sambamurti K, Bennett DA. 2004. Apolipoprotein gene and its interaction with the environmentally driven risk factors: Molecular, genetic and epidemiological studies of Alzheimer's disease. *Neurobiol Aging* 25(5):651-660.
- Lesne S, Docagne F, Gabriel C, Liot G, Lahiri DK, Buee L, Plawinski L, Delacourte A, MacKenzie ET, Buisson A, et al. 2003. Transforming growth factor-beta 1 potentiates amyloid-beta generation in astrocytes and in transgenic mice. *J Biol Chem* 278(20):18408-18418.
- Libby P. 2001. Current concepts of the pathogenesis of the acute coronary syndromes. *Circulation* 104(3):365-372.
- Luedeking EK, DeKosky ST, Mehdi H, Ganguli M, Kamboh MI. 2000. Analysis of genetic polymorphisms in the transforming growth factor-beta1 gene and the risk of Alzheimer's disease. *Hum Genet* 106(5):565-569.
- Lunetta KL, Faraone SV, Biederman J, Laird NM. 2000. Family-based tests of association and linkage that use unaffected sibs, covariates, and interactions. *Am J Hum Genet* 66(2):605-614.
- Mantel N, Haenszel W. 1959. Statistical aspects of the analysis of data from retrospective studies of disease. *J NCI* 22:719-748.
- Markesbery WR. 1999. The role of oxidative stress in Alzheimer disease. *Arch Neurol* 56(12):1449-1452.
- Mattson MP, Barger SW, Furukawa K, Bruce AJ, Wyss-Coray T, Mark RJ, Mucke L. 1997. Cellular signaling roles of TGF beta, TNF alpha and beta APP in brain injury responses and Alzheimer's disease. *Brain Res Rev* 23(1-2):47-61.
- Miyata M, Smith JD. 1996. Apolipoprotein E allele-specific antioxidant activity and effects on cytotoxicity by oxidative insults and beta-amyloid peptides. *Nat Genet* 14(1):55-61.
- Monning U, Sandbrink R, Banati RB, Masters CL, Beyreuther K. 1994. Transforming growth factor beta mediates increase of mature transmembrane amyloid precursor protein in microglial cells. *FEBS Lett* 342(3):267-272.
- Neve RL, McPhie DL, Chen Y. 2000. Alzheimer's disease: A dysfunction of the amyloid precursor protein(1). *Brain Res* 886(1-2):54-66.
- Perry RT, Collins JS, Harrell LE, Acton RT, Go RC. 2001. Investigation of association of 13 polymorphisms in eight genes in southeastern African American Alzheimer disease patients as compared to age-matched controls. *Am J Med Genet* 105(4):332-342.
- Poirier J. 2005. Apolipoprotein E, cholesterol transport and synthesis in sporadic Alzheimer's disease. *Neurobiol Aging* 26(3):355-361.
- Suthanthiran M, Li B, Song JO, Ding R, Sharma VK, Schwartz JE, August P. 2000. Transforming growth factor-beta 1 hyperexpression in African-American hypertensives: A novel mediator of hypertension and/or target organ damage. *Proc Natl Acad Sci USA* 97(7):3479-3784.
- Tierney MC, Fisher RH, Lewis AJ, Zorzitto ML, Snow WG, Reid DW, Nieuwstraten P. 1988. The NINCDS-ADRDA Work Group criteria for the clinical diagnosis of probable Alzheimer's disease: A clinicopathologic study of 57 cases. *Neurology* 38(3):359-364.
- van der Wal EA, Gomez-Pinilla F, Cotman CW. 1993. Transforming growth factor-beta 1 is in plaques in Alzheimer and Down pathologies. *Neuroreport* 4(1):69-72.
- Wang P, Yang G, Mosier DR, Chang P, Zaidi T, Gong YD, Zhao NM, Dominguez B, Lee KF, Gan WB, et al. 2005. Defective neuromuscular synapses in mice lacking amyloid precursor protein (APP) and APP-Like protein 2. *J Neurosci* 25(5):1219-1225.
- Wilmer WA, Dixon CL, Hebert C, Lu L, Rovin BH. 2002. PPAR-alpha ligands inhibit H2O2-mediated activation of transforming growth factor-beta1 in human mesangial cells. *Antioxid Redox Signal* 4(6):877-884.
- Wood NA, Thomson SC, Smith RM, Bidwell JL. 2000. Identification of human TGF-beta1 signal (leader) sequence polymorphisms by PCR-RFLP. *J Immunol Methods* 234(1-2):117-122.
- Wyss-Coray T, Feng L, Masliah E, Ruppe MD, Lee HS, Toggas SM, Rockenstein EM, Mucke L. 1995. Increased central nervous system production of extracellular matrix components and development of hydrocephalus in transgenic mice overexpressing transforming growth factor-beta 1. *Am J Pathol* 147(1):53-67.
- Yamada Y, Miyauchi A, Goto J, Takagi Y, Okuizumi H, Kanematsu M, Hase M, Takai H, Harada A, Ikeda K. 1998. Association of a polymorphism of the transforming growth factor-beta1 gene with genetic susceptibility to osteoporosis in postmenopausal Japanese women. *J Bone Miner Res* 13(10):1569-1576.
- Zetterberg H, Andreasen N, Blennow K. 2004. Increased cerebrospinal fluid levels of transforming growth factor-beta1 in Alzheimer's disease. *Neurosci Lett* 367(2):194-196.